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2	What We Don't Know About the Effects of
3	Temperature on Salmonid Growth
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18	Running head: Temperature and salmonid growth
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### 20 ABSTRACT

21 Salmonid biologists use both mental models-simple assumptions-and bioenergetics-22 based mathematical models to understand and predict effects of temperature regime on growth. 23 Bioenergetics model results, however, sometimes conflict with common assumptions. Previous 24 studies plus "Wisconsin model" bioenergetics simulations lead to four conclusions that conflict with some management assumptions. (1) Food consumption is at least as important as 25 26 temperature in explaining growth; we cannot understand temperature effects without 27 understanding food consumption. (2) At natural food consumption rates, there is no "optimal 28 temperature for growth"; growth peaks in model results are artifacts of food consumption 29 assumptions, and in laboratory studies are (apparently) artifacts of ad libitum feeding. (3) Effects 30 of temperature on growth can be stronger during cooler seasons than in summer; traditional 31 temperature criteria are not useful for managing such effects. (4) Salmonid populations adapted 32 to survive higher temperatures may be more, not less, vulnerable to temperature effects on 33 growth, due to their higher metabolic rates. Temperature-growth relations observed under ad 34 libitum feeding seem risky for managing wild populations. Model predictions of growth need to 35 consider food consumption assumptions carefully. For predicting effects of increased 36 temperature, the traditional assumption that consumption is a constant fraction of maximum 37 consumption rate appears especially uncertain and incautious, with its hidden assumption that 38 consumption increases with temperature; assuming a constant ration is simpler and more 39 cautious. Growth can be predicted more reliably with feeding models and individual-based 40 population models that consider how consumption and energetic costs depend on processes such 41 as habitat selection, competition, and adaptive behaviors that trade off food intake and predation 42 risk. Two research needs are clear: empirical observations for parameterizing and testing the 43 Wisconsin model comprehensively under natural conditions (which we lack despite the extensive 44 energetics literature), and methods for predicting salmonid food production responses to 45 temperature and flow regimes.

# 46 **INTRODUCTION**

The effect of temperature regime on stream salmonids has long been an important
management concern, and its importance is increasing rapidly. Management and research

49 biologists often address temperature effects at scales from local to regional, e.g., how 50 temperature interacts with physical habitat and other variables to drive adaptive behaviors such 51 as when and where to feed (Railsback et al. 2021a), how changes in flow and temperature regime 52 resulting from (e.g.) hydropower licensing and forest harvest (Leach et al. 2012) affect trout 53 abundance, and how climate change interacts with other stressors to affect population viability 54 (Ayllón et al. 2021). These questions usually depend less on acute temperature stress and more 55 on sublethal effects such as how temperature affects growth, because sublethal effects happen 56 over wider and lower ranges of temperature.

57 Salmonid biologists use both mental models—assumptions—and mathematical models to 58 understand and predict effects of temperature on growth. Unfortunately, some widely used 59 assumptions sometimes conflict with the mathematical models, as I explore below. These 60 conflicts indicate that, despite an extensive literature, important uncertainties about how 61 temperature affects salmonid growth persist.

62 The specific mathematical model of temperature effects on growth I refer to is the "Wisconsin" bioenergetic model (Hanson et al. 1997; Deslauriers et al. 2017), which includes a 63 64 conceptual model and specific equations and parameters for various species and life stages. The 65 conceptual model is an energy balance, treating growth rate as the difference between the rates of 66 energy intake from food and energy costs of basal metabolism, activity, and digestion. The 67 conceptual and mathematical models are of course great simplifications of complex 68 physiological mechanisms. For example, Wisconsin model applications typically ignore the 69 energetics of reproduction and adaptive energy allocation (e.g., growth in size vs. storage as 70 lipids).

71 However, bioenergetics modeling is popular as a way to relate fish growth to temperature 72 and other variables because it is simple. Unlike another widely used energetics approach 73 (Dynamic Energy Budget theory; Kooijman 2010; Nisbet et al. 2000), the Wisconsin model has 74 two characteristics making it useful for management modeling. First, it uses only a minimal set 75 of variables to link temperature and growth: growth rate, energy intake, temperature, fish size, 76 activity. Second, the model's energy compartments and mechanisms are clearly related to actual 77 physiology and therefore measurable, at least in principle. We can put fish in the laboratory and 78 measure, e.g., how metabolic energy demand varies with temperature and swimming speed. 79 Therefore, the Wisconsin model provides a framework for combining observations from various

experiments into a predictive tool. It has become a very important tool not just by itself but as the
growth component of numerous models (cited throughout this article) of phenomena from
individual growth to population dynamics to regional productivity.

83 Unfortunately, despite extensive literature on relevant mechanisms, we still lack 84 conclusive empirical evidence for evaluating both mental and mathematical models of 85 temperature effects on salmonid growth. To my knowledge, there has not been a single study on 86 real fish that carefully controlled and independently varied the four drivers of growth in the 87 Wisconsin model (energy intake, temperature, fish size, and swimming speed) while observing 88 growth. Salmonid biologists are blessed with probably hundreds of studies of how some of these 89 variables affect growth, but none that we can use as a thorough and conclusive test of the 90 Wisconsin model.

91 The fundamental problem with the empirical literature is that quantitative results depend 92 on factors that always vary among experiments; examples include species and stock, exercise 93 condition, life history stage, food type, and measurement methods (Hartman and Brandt 1993). 94 Consequently, results from different experiments often conflict (even when experiments use very 95 similar methods and equipment; e.g., Forseth and Jonsson 1994; Myrick and Cech 2000), and 96 combining them into a useful set of equations and parameters requires judgement and 97 assumptions that introduce uncertainty. Other problems with many laboratory studies include the 98 use of ad libitum feeding and artificial food, which are convenient but produce unnaturally food-99 rich, high-energy conditions; not taking observations at the extreme temperatures that are often 100 of management concern, or not observing enough temperatures to define the sharp changes that 101 can occur at high temperatures; and unmeasurable variation in food intake and swimming speed 102 due to competitive interactions among multiple fish kept in the same tank (e.g., Forseth and 103 Jonsson 1994).

104 My objective here is to identify conflicts between management assumptions and models 105 and examine them in light of the available evidence, thereby illuminating key research needs and 106 ways we can improve both mental and mathematical models. I identify four assumptions about 107 how temperature affects growth that I have noted in the literature and in interactions with 108 salmonid biologists. I then examine how well these assumptions agree with the bioenergetic 109 model and, sometimes, empirical evidence. The Discussion examines key conflicts between our

mental and mathematical models, suggests alternative assumptions, and identifies researchobjectives to reduce key uncertainties.

## 112 MANAGEMENT ASSUMPTIONS AND MODEL PREDICTIONS

### 113 Assumption 1: Growth is Driven by Temperature

Perhaps the most fundamental assumption salmonid biologists make about temperature is that it has a strong, even predominant, effect on growth: if we see that trout in one stream are larger than those in another stream, we often assume that the difference results from the first stream being warmer. This assumption also appears in management models. For example, Mims et al. (2019) used temperature as the sole habitat variable driving growth in a large-scale metapopulation model, and Lopez Arriaza et al. (2017) modeled growth in juvenile Steelhead Trout as a function of fish activity, competition, and temperature as the only habitat variable.

121 The Wisconsin model offers the alternative assumption that differences in growth can 122 also be explained by differences in energy intake, either via higher food consumption or higher 123 prey energy density. In fact, a comprehensive parameter sensitivity analysis of three versions of 124 the Wisconsin model (none for salmonids, however) found that parameters affecting food intake 125 had the strongest effect on results (Bartell et al. 1986). Stewart et al. (1983) and Beauchamp et 126 al. (1989) found high sensitivity to food intake parameters in salmonid applications of the model.

127 I examined the relative effects of temperature and food consumption with a simple 128 sensitivity experiment: I executed the Wisconsin model for both temperature and ration ranging 129 from -20% to +20% of baseline values. (Following Deslauriers et al. 2017, I use "ration" as an 130 individual's mass rate of food consumption, g/d; energy intake rate is equal to ration times a prev 131 energy density.) I used Version 1.1.3 of the Fish Bioenergetics Model 4 software (Deslauriers et 132 al. 2017), its standard formulation for adult Rainbow Trout, and its default prey and predator 133 energy densities of 3000 and 4500 J/g. I simulated growth of a 40-g trout over 30 days with 134 constant temperature and ration. The baseline value of ration, 2.06 g/d, was calculated as that 135 producing zero growth at 20°, representing realistic intake in a natural, relatively oligotrophic environment. The baseline temperature was 15°. Therefore, I varied temperature 12–18° with 136 137 ration of 2.06 g/d, then ration 1.65–2.47 g/d with temperature at 15°.

138 The results of this experiment (Figure 1) indicate that food—energy intake rate—affects 139 growth as much or more than temperature does. Food intake seems even more important when 140 we consider that it often varies far more widely than temperature among sites, seasons, and 141 perhaps management scenarios such as alternative flow regimes; a decrease in flow, for example, 142 may produce an increase of several degrees in summer temperature but a large decrease in drift 143 food delivery (e.g., Harvey et al. 2006). Therefore, it appears risky to assume that differences in 144 growth (among sites, over time, etc.) are due to temperature alone and not also to food 145 availability. In fact, we should also consider other factors that affect the energy balance, e.g., 146 differences in activity (e.g., between low- and high-slope sites) or food quality (e.g., between 147 times when food is dominated by immature aquatic vs. adult or terrestrial insects). 148



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Figure 1. Results of the temperature and ration sensitivity experiment for adult rainbow trout. The X axis is ration or temperature, as the percentage change from its baseline value, and the Y axis is predicted fish weight after 30 days.

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### 153 Assumption 2: There is an Optimal Temperature for Growth

The second assumption I examine is that growth typically increases with temperature up to an "optimal" temperature, above which it decreases. A number of studies have tried to quantify optimal growth temperatures (e.g., Forseth and Jonsson 1994; Myrick and Cech 2000), and the assumption has been used in management models (e.g., Fullerton et al. 2017; Lopez Arriaza et al. 2017) and recommendations (e.g., Zeigler et al. 2013), and in other analyses of

159 temperature effects (e.g., Huntsman et al. 2021).

160 I evaluated this assumption simply by running the Wisconsin model for adult Rainbow 161 Trout over a range of temperatures, holding food consumption constant. However, I repeated the 162 experiment using three definitions of food consumption. One is absolute ration, g food per day. Second is relative ration, g food per g fish per day, so absolute ration increases as the fish grows. 163 164 Third is p, a measure peculiar to the Wisconsin model. The value of p is ration as a fraction of 165 the fish's maximum consumption rate (Cmax, g/d); the model represents Cmax as a nonlinear 166 function of both fish weight and temperature (Figure 2). Assuming a constant *p* therefore means 167 that absolute ration increases with weight and varies with temperature; for the adult Rainbow 168 Trout formulation, ration at a constant p increases sharply with temperature up to  $>20^{\circ}$ C. 169



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Figure 2. Temperature functions for *Cmax* and, for comparison, respiration, in the adult Rainbow Trout formulation
 provided with the Fish Bioenergetics 4.0 software of Deslauriers et al. (2017).

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174 As in the first experiment, I selected consumption parameter values representing 175 moderate energy intake, producing zero growth at  $20^{\circ}$ C: absolute ration = 2.06 g/d, relative ration = 0.0515 g·g<sup>-1</sup>·d<sup>-1</sup>, and p = 0.265. I also simulated high energy intake: absolute ration = 176 4.50 g/d, relative ration = 0.0910 g·g<sup>-1</sup>·d<sup>-1</sup>, and p = 0.500. These high intake rates produce 20 g 177 178 of growth over 30 d at 20°C, an average of 1.4% body weight per day; for comparison, Myrick 179 and Cech (2000) observed growth in the range of 3.0-3.7%/d at 19 and 22°C with ad libitum 180 feeding of high-energy pellets. I simulated 30 d of growth for a trout starting at 40 g, at eight 181 constant temperatures from 5 to 22.5°C.

182 The results for moderate energy intake (Figure 3, top panel) show that an "optimal" 183 growth temperature only occurs when we assume food consumption is a constant fraction of 184 *Cmax*, not when we simulate a constant energy intake rate. When the model assumes fish have 185 the same absolute energy intake, and metabolic energy demands increase with temperature (up to 186 a peak at 22°C, in this case; Figure 2), then growth rate can only decline as temperature 187 increases. However, when the model assumes that food consumption follows the complex 188 function for *Cmax*, which also increases with temperature, then a peak in growth as temperature 189 increases is possible as an artifact of the *Cmax* temperature function. (Using similar methods, 190 Beachamp 2009 showed that the temperature producing highest predicted growth depends on p.)

With high energy intake (Figure 3, bottom panel), there are peaks in growth at relatively low temperatures when intake is modeled as a constant absolute or relative ration. These peaks occur because the Wisconsin model limits the food consumption rate to the value of *Cmax*, which at low temperatures is lower than the specified absolute and relative rations; the simulated fish actually gets less food than the assumed intake rates.

196 If this simulation experiment indicates that peaks in simulated growth with temperature 197 are artifacts of assumptions about Cmax, why have "optimal" temperatures been found in 198 laboratory studies of real salmonids? One answer appears to be that such studies typically use ad 199 libitum feeding (e.g., Zeigler et al. 2013; Myrick and Cech 2000; Forseth and Jonsson 1994) so 200 growth is limited by digestion rate instead of the balance between a limited energy intake and 201 metabolic costs that increase with temperature. (In fish such as grazers that use less-digestible 202 food, temperature appears to have strong effect on growth because digestion rates are strongly 203 temperature-dependent, and energy assimilation is typically limited by digestion instead of intake 204 rates; e.g., Hofer et al. 1982.) Slower digestion at lower temperatures is likely what causes the 205 appearance of an optimal growth temperature in experiments with ad libitum feeding (and the 206 reason *Cmax* is low at low temperatures). However, at more natural food consumption rates the 207 energy assimilation of salmonids is unlikely to be limited by digestion rate, except perhaps at 208 low temperatures. There is some evidence to support this potential explanation. Wurtsbaugh and 209 Davis (1977) observed growth in juvenile Rainbow Trout under three temperature regimes and 210 several ration levels. At low rations, growth always decreased with temperature, but temperature 211 had positive effects on growth at higher rations. Cui and Wootton (1988) observed growth of 212 minnows fed on small invertebrates at five ration levels and four temperatures; they too found

growth to only decrease with temperature at low rations, but peak or increase with temperature at high rations. The concept of an optimal temperature for growth may be useful in situations where food is essentially unlimited and energy-rich-hatcheries and prey pulses (Furey et al. 2016)-but appears questionable for salmonids feeding on invertebrate prey, in which case growth is much more likely to be limited by food capture rate than digestion rate. 



(as in Figure 1) for three food consumption assumptions. Top panel: moderate food intake producing zero growth at 20°C. Bottom panel: high intake that produces 1.4%d<sup>-1</sup> growth at 20°C.

This experiment clearly shows that the response of growth to temperature depends, even qualitatively, on what happens to food consumption. The differences among ration assumptions in Figure 3 have important management implications. For example, when we assume absolute ration is constant, then growth under moderate energy intake is most sensitive to temperature at

intermediate temperatures,  $\sim 7-17^{\circ}$ ; when we assume constant *p*, then growth is insensitive to temperature at  $\sim 10-17^{\circ}$  and most sensitive above and below that range. Management recommendations based on temperature-growth relations observed under ab libitum feeding (e.g., Zeigler et al. 2013) seem especially risky.

#### **Assumption 3: Temperatures Effects are Important Only in Summer**

We often naturally assume that the highest summer temperatures are our biggest management concern for salmonids. This assumption is reflected in the widespread use of temperature criteria for assessing temperature effects of management alternatives (e.g., by evaluating how frequently a criterion of 20°C is exceeded under each alternative). Peak temperatures are an obvious concern in streams where natural climate and human impacts drive them to levels making growth difficult and acute effects likely (e.g., Ayllón et al. 2013).

240 Studies applying bioenergetics models to year-round growth at sites with less-extreme 241 thermal regimes, however, have found that temperature can have strong effects on growth during 242 seasons other than summer. Railsback and Rose (1999) analyzed temperature effects on growth 243 using the Wisconsin model and trout sizes observed in spring and fall at a number of sites in the 244 Sierra Nevada, California. They concluded that temperature had stronger effects from fall to 245 spring than from spring to fall. Similarly, Armstrong et al. (2021) applied the model of Fullerton 246 et al. (2017), which also uses a Wisconsin model formulation to represent growth, to year-round 247 temperatures throughout several watersheds in the Pacific Northwest. They concluded that 248 lower-elevation sites offering low summer growth due to higher temperatures provided important 249 high-growth habitat at other times of year.

These analyses illustrate that changes in temperature regime can have strong effects on salmonid growth during cooler seasons. Figure 3 illustrates why the Wisconsin model predicts this: for all the ration assumptions, the sensitivity of growth to temperature is high at low to intermediate temperatures.

### **Assumption 4: Warm-adapted Populations Grow Better at Higher Temperatures**

Clear evidence shows that some salmonid populations are better adapted to high
temperatures than others, even within a species (e.g., Eliason et al. 2011; Chen et al. 2015, 2018).
This evidence is mostly derived from measurements of acute thermal effects such as critical

thermal maximum (the temperature at which a fish loses swimming ability during rapidly

259 increasing temperatures) and aerobic scope (the difference between resting and maximum

260 metabolic rates at elevated temperatures, which reflects how resting metabolic rate increases and 261 maximum rate decreases with temperature; e.g., Eliason et al. 2011). Larger hearts and increased 262 cardiac performance appear to be key mechanisms conveying acute temperature tolerance (Chen 263 et al. 2018).

264 We might naturally assume that warm-adapted salmonid populations have lower 265 sublethal effects: that populations evolved to tolerate higher temperature extremes are also likely 266 to grow over a wider range of temperatures. However, the mechanism of adaptation—increased 267 heart size and pumping rate—seems likely to come at a cost of higher metabolic rate. In fact, 268 recent evidence indicates the validity of this concern. Data from two southern California 269 Steelhead populations indicate that the population from a warmer stream has higher tolerance of 270 extreme temperatures and approximately double the resting metabolic rate, measured over 271 temperatures of 18–21°C (Eliason, pers. comm.). It seems reasonable to assume that this 272 difference in resting metabolic rate would extend to lower temperatures as well.

If higher tolerance of extreme temperature has a cost of higher metabolic rates, then we should not think of "warm-adapted" populations (and, perhaps, species) as less subject to effects of temperature on growth. Just the opposite appears to be true: such fish may be able to grow and thrive only when able to consume considerably more food than others, to keep up with their higher metabolic costs. This evidence also implies that we should reconsider Wisconsin model respiration parameters to avoid overestimation of growth, or underestimation of food requirements, for warm-adapted populations.

#### 280 **DISCUSSION**

#### 281 Conflicts Between Management Assumptions and Bioenergetics Modeling

The Wisconsin bioenergetics model has proven extremely useful as a temperature management tool. It is useful not because it is "realistic"—it is not—but, in contrast, because it is as simple as possible while still linking the management variables we need to evaluate, via mechanisms we can measure. The model provides a way to assemble relations observed in diverse laboratory studies into a predictive tool. My comparison of Wisconsin model results to management assumptions leads to the following conclusions, some of which challenge how wecommonly think about temperature effects.

Growth is driven by food as much or more than by temperature. When trying to explain differences or changes in growth, we should consider food consumption, not temperature, as the primary driver. Further, there is no way to understand the effects of temperature without understanding food consumption; Figure 3 shows that even the shape of the growth response to temperature depends on what we assume about food consumption.

294 There may be no optimal temperature for growth under typical natural conditions. I 295 found a peak in simulated growth rate with temperature only as an artifact of assuming ration 296 varies with *Cmax*, not under simpler and perhaps more realistic consumption assumptions. 297 Optimal growth temperatures found in laboratory studies seem likely an artifact of ad libitum 298 feeding, which makes growth limited by digestion rate instead of food intake rate. While there 299 may be situations (including low temperatures) when wild salmonids can capture food more 300 rapidly than they can digest it, under typical natural feeding conditions (e.g., drift-feeding on 301 invertebrates), salmonid growth is much more likely limited by food intake than by digestion. 302 Therefore, relations between growth and temperature observed under unnaturally food-rich 303 laboratory conditions seem a risky basis for management of wild populations.

304 Peak temperatures may not be the most important to manage. Temperature can have
 305 strong effects on growth during cool seasons, which implies that traditional temperature criteria
 306 have limited value for assessing and avoiding growth effects.

Warm-adapted salmonid populations may be more, not less, vulnerable to
temperature effects on growth. The higher metabolic rates that allow survival of higher
extreme temperatures in some populations also require higher food intake to maintain growth.
Consequently, populations that have evolved tolerance of higher temperatures must be assumed
more vulnerable to elevated temperatures that are not accompanied by higher food availability.

### 312 **Representing Consumption in Bioenergetics Analyses**

The main lesson from my analyses is that we should not ignore food in modeling how temperature affects salmonid growth. Unfortunately, while temperature is easy to measure and model, food availability and consumption by fish are complex, expensive to observe, and impossible to predict with certainty. Therefore, we almost always rely on simple assumptions about food; typically, we calibrate a consumption parameter by fitting the model to growth
observed under one temperature regime and then use that parameter value to predict growth
under other temperature regimes.

What is a good assumption about food consumption for predicting growth under alternative temperatures? If management conclusions drawn from analyses based on the Wisconsin model are likely to change if we assume different consumption rates, can those analyses have any credibility? These questions are especially important when examining management actions, e.g., changes in flow, that are likely to affect food production as well as temperature.

326 The Wisconsin model literature and software long promoted the practice of representing 327 consumption as a constant value of p, an assumption that seems especially risky and difficult to 328 justify. That assumption causes absolute ration and energy intake rate to vary with temperature 329 according to the complex relation assumed between Cmax and temperature (Figure 2). This 330 variation makes it harder to interpret results and understand exactly what drove predicted growth (Railsback and Rose 1999). Because Cmax typically increases with temperature except at high 331 332 temperatures, assuming constant *p* includes a hidden assumption that ration increases with 333 temperature; therefore, it could underestimate effects of increased temperature on growth (but 334 see below concerning temperature effects on ration). Other problems with this assumption 335 include the lack of an ecological reason why a fish would or could adjust its consumption to 336 maintain constant p as temperature changes (I discuss behavioral aspects of consumption below), 337 and that *Cmax* is a particularly challenging variable to define precisely and measure (Hartman 338 and Brandt 1993)—it can vary with factors that are rarely controlled and not included in the 339 model, such as exercise condition of the fish, food type, activity, and measurement time scale. 340 Figure 2 makes it clear that uncertainty in the Rainbow Trout Cmax temperature function has 341 especially strong effects on predicted growth at temperatures  $>20^{\circ}$ C. (This uncertainty is why I 342 simulated growth at temperatures no higher than 22.5°.) Further, the physiological mechanism 343 driving *Cmax*—digestion rate—is unlikely to limit growth under consumption rates typical of 344 natural conditions (except, possibly, when temperatures are low and food availability high). 345 The alternative assumption of constant ration (either absolute or relative) is simpler and 346 more conservative, by not including the hidden assumption that ration increases with

347 temperature. Calibrating the Wisconsin model to estimate a constant ration that is then used to

estimate effects of other temperature regimes seems simple and clear, and cautious for examining
temperatures above those used for calibration. As an example, Hawkins et al. (2020) modified
their previous model that assumed a constant *p* (Fullerton et al. 2017) to instead use a constant
relative ration.

352 On the other hand, assuming a constant ration ignores how the productivity of aquatic 353 ecosystems actually can vary with temperature; it is not unreasonable to expect a positive 354 relationship between salmonid food intake and temperature. Filbert and Hawkins (1995) 355 observed drift food concentrations roughly an order of magnitude higher in summer than winter, 356 in a reservoir tailwater. From a meta-analysis of published benthic insect growth rates, Morin 357 and Dumont (1994) produced a regression model of growth rate vs. temperature for a 358 combination of Diptera, Ephemeroptera, Plecoptera, and Trichoptera, observed over 359 temperatures from <5 to >20°C. This model predicts insect growth to increase as an exponential function of temperature (growth is proportional to  $e^{0.058T}$  where T is temperature, °C). Neglecting 360 361 complexities such as how insect growth translates to drift rate and how drift depletion by fish 362 varies with temperature, we can use this regression model as a speculative example of how food 363 availability could vary with temperature.

364 To explore the speculation that trout food consumption increases with temperature 365 according to the growth equation of Morin and Dumont (1994), I repeated the simulation 366 experiment of Figure 3 while assuming that food consumption varies, above and below 15°C, according to their exponential function. I used an absolute ration (g/d) equal to  $0.87e^{0.058T}$ , which 367 368 at 15°C produces the same ration of 2.06 g/d assumed in the "Absolute ration" results of Figure 3 369 (top panel). The results (Figure 4) are completely different from those of the other consumption 370 assumptions: growth is insensitive to temperature up to 17.5°, then *increases* with temperature 371 up to 22.5°.

This simulation provides only a very simplistic view of potential effects of food availability increasing with temperature and should not be used to inform management decisions; however, it further confirms the importance of food consumption assumptions when predicting temperature effects on growth, and to some extent even brings into question the generalization that increased temperature always has negative effects on salmonid growth and population biomass accumulation. It raises the possibility that increased benthic productivity could largely offset the increased metabolic demands of fish; as Railsback and Rose (1999) suggest, the 379 strongest effect of temperature on trout growth could be an indirect effect via food production.

380 Such increased productivity could help explain the persistence of warm-adapted populations

381 despite their higher metabolic rates.

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Figure 4. Growth simulations as in the top panel of Figure 3, with the additional assumption that food consumption follows the temperature term of the insect growth equation of Morin and Dumont (1994).

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387 Unfortunately, even the range of assumptions illustrated in Figure 4 is a gross 388 simplification of food consumption. The consumption rate of each individual fish in a population 389 can also depend on competition for food, and on behavior. Feeding behavior-where an 390 individual feeds, how often, and at what times of day—is now understood and modeled as an 391 adaptive tradeoff between meeting energy demands and avoiding predation risk (Railsback et al. 392 2020). This understanding suggests that the most reasonable simple assumption about food 393 consumption is that each individual feeds enough to meet its metabolic demands and, during 394 relevant life stages, to accumulate the size and energy needed for future survival and 395 reproduction (e.g., Biro et al. 2005). Consequently, the effects of temperature could manifest as 396 changes not in growth but in survival: if increased temperatures raise metabolic rates but not 397 prey availability, salmonids are likely to respond by feeding more often, or in more productive 398 but riskier places and times, and hence to maintain their growth but experience more predation. 399 Lower temperature or higher food availability could result in less time spent feeding and higher 400 survival, instead of (or in addition to) higher growth.

Given all these complexities in estimating consumption, is it possible to make useful and credible predictions of growth (and survival) responses to temperature? At a minimum, we can try several simple food assumptions and see whether and how the conclusions of management studies differ among them (e.g., the food availability experiment of Railsback et al. 2021a).

Incorporating the Wisconsin formulation within individual feeding models and
individual-based population models (IBMs) is a second way to deal with food consumption
complexities. Models that predict individual food intake from habitat conditions have a long
history (e.g., Fausch 1984; Hughes and Dill 1990; Naman et al. 2020). Fullerton et al. (2017),
Mims et al. (2019), and Railsback et al. (2021a, b) provide examples of IBMs, for different
purposes, that combine the Wisconsin formulation with models of how habitat, competition, and
behavior affect food consumption.

412 InSTREAM and inSALMO (Railsback and Harvey 2002; Railsback et al. in prep., 413 2021b) are unique as salmonid population models that represent individual growth (and 414 population abundance and biomass) as outcomes of prey production, physical habitat, 415 temperature, competition, and behaviors that determine when and where individuals feed as a 416 tradeoff between growth and predation risk. These two models are especially valuable for 417 assessing changes in temperature that accompany changes in flow: they predict the combined 418 effects of flow (e.g., on food delivery and habitat space) and temperature on populations 419 (Railsback et al. 2021a, b). These models also facilitate experiments to examine sensitivity of 420 results to alternative assumptions about food; for example, Railsback et al. (2021a) showed that 421 conclusions about a range of instream flow and temperature management alternatives changed 422 little when drift food availability was assumed concentrated during crepuscular periods instead of 423 constant throughout the day. (InSTREAM and inSALMO have the additional benefit of 424 representing temperature effects other than on growth; the application of Railsback et al. 2021b 425 found temperature effects on egg mortality and incubation rate to clearly affect abundance.) 426 These models that predict food consumption from foraging behavior depend on a part of

427 the Wisconsin model otherwise neglected here: the energy cost of swimming. The models treat 428 selection of drift-feeding sites as a tradeoff between the food delivery benefits of higher velocity 429 and its costs in reduced capture ability and increased activity respiration. (In developing the 430 newest version of inSTREAM, we discovered an important flaw in the Wisconsin model's 431 activity respiration formulation, which it adopts from Stewart et al. 1983. It treats activity

- 432 respiration as a function only of swimming speed, neglecting the effect of fish size. See
- 433 Railsback et al. in prep. for an alternative formulation, plus reviews of other components of the
- 434 Wisconsin salmonid formulations in light of recent empirical data.)

#### 435 **Research Needs**

436 As a simple representation of complex physiological and ecological processes, the 437 Wisconsin model depends on empirical data for parameterization and validation; yet we lack 438 even a single comprehensive data set for salmonids, as widely studied and important as trout and 439 salmon are. While a few parameterizations have been based mostly on a single set of laboratory 440 experiments designed specifically for that purpose (e.g., Mesa et al. 2013 for Bull Trout feeding 441 on fish), or shown to predict growth well under limited conditions (Mesa et al. 2013; Madenjian 442 et al. 2004, for Chinook Salmon feeding on fish), many (including the formulation for Rainbow 443 Trout used here) have been cobbled together from multiple studies of questionable compatibility 444 (Railsback and Rose 1999) and, in retrospect, limited value for formulating or testing the model. 445 In fact, the Rainbow Trout formulation produces implausible results above 22.5°C, a critical 446 range for temperature assessment, clear evidence that it needs improvement.

447 To make results reliable and credible, we need laboratory experiments designed 448 specifically to parameterize and test the Wisconsin model as applied to natural conditions. 449 Traditionally measured variables such as Cmax and temperature "tolerance" are not useful for 450 this purpose. Instead, we need observed growth of individual fish of various sizes, exposed to 451 many combinations of temperature, energy intake rate, and swimming speed over wide ranges of 452 these variables (including extremes), in experiments that carefully control the many other factors 453 that affect growth. Data on how metabolic rates of the same fish vary with temperature and 454 swimming speed would help parameterize those components of the model and test the overall 455 energy balance assumptions. Such experiments are likely to be challenging and expensive, yet 456 well within the capabilities of existing laboratory technology.

The evidence that growth depends as much or more on food consumption as on temperature indicates that we also need useful observations and models of how production of salmonid food depends on the variables we predict growth responses to, especially flow and temperature regimes. IBMs that predict how population-level abundance and biomass depend on temperature regime, considering complexities such as competition and behavior, still depend on 462 assumptions about food production rates. Even though salmonid food production is notoriously

- 463 variable and uncertain, additional studies to observe it and learn how it varies should improve
- 464 our ability to predict temperature effects. To improve our ability to predict how temperature (and
- 465 flow) regimes affect salmonids, we do not need a comprehensive understanding of food
- 466 production dynamics; instead, we need to understand those dynamics just well enough to select
- 467 simple yet valid assumptions of how food availability varies among those regimes.

#### 468 **REFERENCES**

- 469 Armstrong, J. B., A. H. Fullerton, C. E. Jordan, J. L. Ebersole, J. R. Bellmore, I. Arismendi, B.
- 470 E. Penaluna, and G. H. Reeves. 2021. The importance of warm habitat to the growth regime
- 471 of cold-water fishes. Nature Climate Change 11:354–361.
- 472 Ayllón, D., G. G. Nicola, B. Elvira, and A. Almodóvar. 2021. Climate change will render size-
- 473 selective harvest of cold-water fish species unsustainable in Mediterranean freshwaters.
  474 Journal of Applied Ecology 58:562–575.
- 475 Ayllón, D., G. G. Nicola, B. Elvira, I. Parra, and A. Almodóvar. 2013. Thermal carrying capacity
  476 for a thermally-sensitive species at the warmest edge of its range. PLoS ONE 8:e81354.
- 477 Bartell, S. M., J. M. Breck, R. H. Gardner, and A. L. Brenkert. 1986. Individual parameter
- 478 perturbation and error analysis of fish bioenergetics models. Canadian Journal of Fisheries
  479 and Aquatic Sciences 43:160–168.
- Beauchamp, D. A., D. J. Stewart, and G. L. Thomas. 1989. Corroboration of a bioenergetics
  model for Sockeye Salmon. Transactions of the American Fisheries Society 118:597–607.
- 482 Beauchamp, D. A. 2009. Bioenergetic ontogeny: linking climate and mass-specific feeding to
- 483 life-cycle growth and survival of salmon. Pages 53–72 in C. Zimmerman and C. C. Krueger,
- 484 editors. Pacific salmon: ecology and management of western Alaska's populations. American
  485 Fisheries Society, Symposium 70, Bethesda, Maryland.
- 486 Biro, P. A., J. R. Post, and M. V. Abrahams. 2005. Ontogeny of energy allocation reveals
- 487 selective pressure promoting risk-taking behaviour in young fish cohorts. Proceedings of the
  488 Royal Society B 272:1443–1448.
- 489 Chen, Z., A. P. Farrell, A. Matala, N. Hoffman, and S. R. Narum. 2018. Physiological and
- 490 genomic signatures of evolutionary thermal adaptation in Redband Trout from extreme
- 491 climates. Evolutionary Applications 11:1686–1699.

- 492 Chen, Z., M. Snow, C. S. Lawrence, A. R. Church, S. R. Narum, R. H. Devlin, and A. P. Farrell.
- 493 2015. Selection for upper thermal tolerance in Rainbow Trout (*Oncorhynchus mykiss*494 Walbaum). Journal of Experimental Biology 218:803–812.
- 495 Cui, Y. and R. J. Wootton. 1988. Bioenergetics of growth of a cyprinid, *Phoxinus phoxinus* (L.):
- 496 the effect of ration and temperature on growth rate and efficiency. Journal of Fish Biology497 33:763–773.
- 498 Deslauriers, D., S. R. Chipps, J. E. Breck, J. A. Rice, and C. P. Madenjian. 2017. Fish
  499 Bioenergetics 4.0: an R-based modeling application. Fisheries 42:586–596.
- Eliason, E. J. Personnel communication. E. J. Eliason, Department of Ecology, Evolution, and
   Marine Biology, University of California, Santa Barbara CA. Email to S. F. Railsback, 11
- 502 June 2021.
- 503 Eliason, E. J., T. D. Clark, M. J. Hague, L. M. Hanson, Z. S. Gallagher, K. M. Jeffries, M. K.
- Gale, D. A. Patterson, S. G. Hinch, and A. P. Farrell. 2011. Differences in thermal tolerance
  among Sockeye Salmon populations. Science 332:109–112.
- Fausch, K. D. 1984. Profitable stream positions for salmonids: relating specific growth rate to net
  energy gain. Canadian Journal of Zoology 62:441–451.
- Filbert, R. B. and C. P. Hawkins. 1995. Variation in condition of Rainbow Trout in relation to
  food, temperature, and individual length in the Green River, Utah. Transactions of the
  American Fisheries Society 124:824–835.
- Forseth, T. and B. Jonsson. 1994. The growth and food ration of piscivorous Brown Trout
  (*Salmo trutta*). Functional Ecology 8:171–177.
- 513 Fullerton, A. H., B. J. Burke, J. J. Lawler, C. E. Torgersen, J. L. Ebersole, and S. G. Leibowitz.
- 514 2017. Simulated juvenile salmon growth and phenology respond to altered thermal regimes515 and stream network shape. Ecosphere 8:e02052.
- 516 Furey, N.B., S.G. Hinch, M.G. Mesa, and D.A. Beauchamp. 2016. Piscivorous fish exhibit
- 517 temperature-influenced binge feeding during an annual prey pulse. Journal of Animal
  518 Ecology 85:1307–1317.
- 519 Hanson, P., T. Johnson, J. Kitchell, and D. E. Schindler. 1997. Fish Bioenergetics 3.0. University
- 520 of Wisconsin Sea Grant Institute, Madison, Wisconsin.

- Hartman, K. J. and S. B. Brandt. 1993. Systematic sources of bias in a bioenergetics model:
  examples for age-0 Striped Bass. Transactions of the American Fisheries Society 122:912–
  926.
- 524 Harvey, B. C., R. J. Nakamoto, and J. L. White. 2006. Reduced streamflow lowers dry-season
- growth of Rainbow Trout in a small stream. Transactions of the American Fisheries Society135:990–1005.
- Hawkins, B. L., A. H. Fullerton, B. L. Sanderson, and E. A. Steel. 2020. Individual-based
  simulations suggest mixed impacts of warmer temperatures and a nonnative predator on
  Chinook Salmon. Ecosphere 11:e03218.
- Hofer, R., H. Forstner, and R. Rettenwander. 1982. Duration of gut passage and its dependence
  on temperature and food consumption in roach, *Rutilus rutilus* L: laboratory and field
  experiments. Journal of Fish Bioliology 20:289–299.
- 533 Hughes, N. F. and L. M. Dill. 1990. Position choice by drift-feeding salmonids: model and test
- for Arctic Grayling (*Thymallus arcticus*) in subarctic mountain streams, interior Alaska.
  Canadian Journal of Fisheries and Aquatic Sciences 47:2039–2048.
- 536 Huntsman, B.M., A.J. Lynch, and C.A. Caldwell. 2021. Interacting effects of density-dependent
- and density-independent factors on growth rates in southwestern Cutthroat Trout populations.
  Transactions of the American Fisheries Society 150: 651-664.
- Kooijman, S. A. L. M. 2010. Dynamic energy budget theory for metabolic organisation.
  Cambridge University Press.
- 541 Leach, J. A., R. D. Moore, S. G. Hinch, and T. Gomi. 2012. Estimation of forest havesting-
- induced stream temperature changes and bioenergetic consequences for Cutthroat Trout in a
  coastal stream in British Columbia, Canada. Aquatic Sciences 74:427–441.
- 544 Lopez Arriaza, J., D. A. Boughton, K. Urquhart, and M. Mangel. 2017. Size-conditional
- smolting and the response of Carmel River Steelhead to two decades of conservation efforts.
  PloS ONE 12:e0188971.
- 547 Madenjian, C. P., D. V. O'Connor, S. M. Chernyak, R. R. Rediske, and J. P. O'Keefe. 2004.
- 548 Evaluation of a Chinook Salmon (*Oncorhynchus tshawytscha*) bioenergetics model.
- 549 Canadian Journal of Fisheries and Aquatic Sciences 61:627–635.

- 550 Mesa, M. G., L. K. Weiland, H. E. Christiansen, S. T. Sauter, and D. A. Beauchamp. 2013.
- 551 Development and evaluation of a bioenergetics model for Bull Trout. Transactions of the
   552 American Fisheries Society 142:41–49.
- 553 Mims, M. C., C. C. Day, J. J. Burkhart, M. R. Fuller, J. Hinkle, A. Bearlin, J. B. Dunham, P. W.

554 DeHaan, Z. A. Holden, and E. E. Landguth. 2019. Simulating demography, genetics, and 555 spatially explicit processes to inform reintroduction of a threatened char. Ecosphere

556 10:e02589.

- Morin, A. and P. Dumont. 1994. A simple model to estimate growth rate of lotic insect larvae
  and its value for estimating population and community production. Journal of the North
  American Benthological Society 13:357–367.
- Myrick, C. A. and J. J. Cech, Jr. 2000. Temperature influences on California Rainbow Trout
  physiological performance. Fish Physiology and Biochemistry 22:245–254.
- 562 Naman, S. M., J. S. Rosenfeld, J. R. Neuswanger, E. C. Enders, J. W. Hayes, E. O. Goodwin, I.
- G. Jowett, and B. C. Eaton. 2020. Bioenergetic habitat suitability curves for instream flow
  modeling: introducing user-friendly software and its potential applications. Fisheries 45:605–
  613.
- Nisbet, R. M., E. B. Muller, K. Lika, and S. A. L. M. Kooijman. 2000. From molecules to
  ecosystems through dynamic energy budget models. Journal of Animal Ecology 69:913–926.
- 568 Railsback, S. F., B. C. Harvey, and D. Ayllón. 2020. Contingent tradeoff decisions with
- feedbacks in cyclical environments: testing alternative theories. Behavioral Ecology
  31:1192–1206.
- 571 Railsback, S. F., B. C. Harvey, and D. Ayllón. 2021a. Importance of the daily light cycle in
  572 population-habitat relations: a simulation study. Transactions of the American Fisheries
  573 Society 150:130–143.
- Railsback, S. F., B. C. Harvey, and D. Ayllón. 2021b. InSTREAM 7: Instream flow assessment
  and management model for stream trout. River Research and Applications. DOI:
  10.1002/rra.3845.
- 577 Railsback, S. F., B. C. Harvey, and D. Ayllón. In preparation. InSTREAM 7 user manual: model

578 description, software guide, and application guide. USDA Forest Service, Pacific Southwest

- 579 Research Station, Albany, California. Available at: https://ecomodel.humboldt.edu/instream-
- 580 7-and-insalmo-7.

- Railsback, S. F. and B. C. Harvey. 2002. Analysis of habitat selection rules using an individualbased model. Ecology 83:1817–1830.
- Railsback, S. F. and K. A. Rose. 1999. Bioenergetics modeling of stream trout growth:
  temperature and food consumption effects. Transactions of the American Fisheries Society
- 585 128:241–256.
- 586 Stewart, D. J., D. Weininger, D. V. Rottiers, and T. A. Edsall. 1983. An energetics model for
- 587 Lake Trout, *Salvelinus namaycush*: application to the Lake Michigan population. Canadian
  588 Journal of Fisheries and Aquatic Sciences 40:681–698.
- 589 Wurtsbaugh, W. A. and G. E. Davis. 1977. Effects of temperature and ration level on the growth
- and food conversion efficiency of *Salmo gairdneri*, Richardson. Journal of Fish Biology
- 591 11:87–98.
- 592 Zeigler, M. P., S. F. Brinkman, C. A. Caldwell, A. S. Todd, M. S. Recsetar, and S. A. Bonar.

5932013. Upper thermal tolerances of Rio Grande Cutthroat Trout under constant and fluctuating

temperatures. Transactions of the American Fisheries Society 142:1395–1405.